

DISC1 is associated with prefrontal cortical gray matter and positive symptoms in schizophrenia

Philip R. Szeszko^{a,b,c,*}, Colin A. Hodgkinson^d, Delbert G. Robinson^{a,b,c},
Pamela DeRosse^b, Robert M. Bilder^e, Todd Lencz^{a,b,c},
Katherine E. Burdick^{a,b,c}, Barbara Napolitano^{a,b}, Julia D. Betensky^b,
John M. Kane^{a,b,c}, David Goldman^d, Anil K. Malhotra^{a,b,c}

^a *Feinstein Institute for Medical Research, Manhasset, NY, USA*

^b *Department of Psychiatry Research, Zucker Hillside Hospital, North-Shore - Long Island Jewish Health System, 75–59 263rd Street, Glen Oaks, NY 11004, USA*

^c *Department of Psychiatry, Albert Einstein College of Medicine, Bronx, NY, USA*

^d *Section on Human Neurogenetics, National Institute on Alcohol Abuse and Alcoholism, Rockville, MD, USA*

^e *UCLA Neuropsychiatric Institute and Geffen School of Medicine, Los Angeles, CA, USA*

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Abstract

Background: *DISC1* is considered a susceptibility gene for schizophrenia and schizoaffective disorder, but little is known regarding the potential mechanisms through which it may confer increased risk. Given that *DISC1* plays a role in cerebral cortex development, polymorphisms in this gene may have relevance for neurobiological models of schizophrenia that have implicated cortical deficits in its pathophysiology.

Methods: We investigated whether the *DISC1* leu607phe polymorphism was associated with prefrontal gray matter volumes using magnetic resonance imaging in a cohort of patients with schizophrenia ($N = 19$) and healthy volunteers ($N = 25$) and positive and negative symptoms in 200 patients with schizophrenia.

Results: Among patients and healthy volunteers, phe carriers ($N = 11$) had significantly less gray matter in the superior frontal gyrus and anterior cingulate gyrus compared to leu/leu homozygotes ($N = 33$). Further, among patients left superior frontal gyrus gray matter volume was significantly negatively correlated with severity of hallucinations. In addition, patients who were phe carriers ($N = 144$) had significantly greater severity of positive symptoms (hallucinations) compared to patients who were leu/leu homozygotes ($N = 56$).

Discussion: These findings implicate *DISC1* in variation of prefrontal cortical volume and positive symptoms, thus providing a potential mechanism through which *DISC1* may confer increased risk for schizophrenia or schizoaffective disorder.

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1. Introduction

There is now considerable evidence that the Disrupted in Schizophrenia 1 (*DISC1*) gene plays a role in schizophrenia (Porteous et al., 2006). *DISC1* is located at the breakpoint of a chromosomal translocation in the 1q42 region resulting in a balanced translocation that cosegregated with schizophrenia and

other affective disorders (St Clair et al., 1990). Subsequent studies indicated that *DISC1* and Disrupted in Schizophrenia II (*DISCII*) were disrupted by the translocation (Millar et al., 2000, 2001; Blackwood et al., 2001). Association studies provided further support for the hypothesis that *DISC1* is a susceptibility gene for schizophrenia and schizoaffective disorder (e.g., Hennah et al., 2003; Cannon et al., 2005). Moreover, in one of the strongest association studies to date our group reported that a missense mutation at a single nucleotide polymorphism (SNP) resulting in a phenylalanine substitution for leucine at position 607 (leu607phe) was overrepresented in patients compared to healthy individuals (Hodgkinson et al., 2004).

* Corresponding author at: The Zucker Hillside Hospital, Department of Psychiatry Research, 75–59 263rd Street, Glen Oaks, NY 11004, USA.
Tel.: +1 718 470 8489; fax: +1 718 343 1659.

E-mail address: szeszko@lij.edu (P.R. Szeszko).

Little is known regarding the mechanisms through which *DISC1* may confer increased risk for schizophrenia. Although the functions of *DISC1* have not been fully elaborated, it may be directly relevant to basic neurodevelopmental mechanisms in the central nervous system that have relevance to the pathogenesis of schizophrenia. For example, *DISC1* has been implicated in brain development (Schurov et al 2004; Kamiya et al 2005), which fits well with neurodevelopmental models of schizophrenia (Bilder and Degreef, 1991; Szeszko et al., 2003). Particularly relevant are data from Kamiya et al. (2005) who reported that suppression of *DISC1* may impair migration and subsequent development of the cerebral cortex.

The identification of endophenotypes in genetics studies may help elucidate the relationship between risk genes such as *DISC1* and phenomenology (Blackwood and Muir, 2004; Szeszko et al., 2005). Surprisingly, however, few studies have investigated relationships among genetic variation in *DISC1*, gray matter structure in vivo and symptom severity in schizophrenia. Some work suggests that genetic variation in *DISC1* may be associated with delusions and hallucinations (Hennah et al., 2003; DeRosse et al., 2007) and that carriers of a 3 locus haplotype, which includes the leu607phe polymorphism, have prefrontal gray matter deficits compared to noncarriers (Cannon et al., 2005). Better understanding how genetic variation in *DISC1* is associated with cortical measures and symptoms in schizophrenia as well as the potential relationship between gray matter structure and symptoms could provide important clues to the neurobiology of schizophrenia.

We tested the hypothesis that the *DISC1* leu607phe polymorphism would be associated with prefrontal gray matter volume and symptoms in patients. We focused on prefrontal regions given that twin studies of schizophrenia suggest that the genetic underpinnings of brain structural deficits may lie in these regions (Cannon et al., 2002). Moreover, both positive (Lennox et al., 2000; Erkwow et al., 1997) and negative symptoms (Sanfilipo et al., 2000) have been linked with prefrontal cortical abnormalities in schizophrenia. Based on prior work (Hennah et al., 2003; Cannon et al., 2005; Hodgkinson et al., 2004) we hypothesized that phe carriers would have less prefrontal cortical gray matter and greater severity of symptoms compared to leu/leu homozygotes. We focused on the *DISC1* leu607phe SNP because it: (a) was most strongly associated with disease in our prior study (Hodgkinson et al., 2004); and (b) was part of a 3 locus haplotype within *DISC1* that was overtransmitted to patients (Hennah et al., 2003; Cannon et al., 2005), which was associated with lower prefrontal gray matter density (Cannon et al., 2005).

2. Method

2.1. Subjects

Twenty-five (11 males, 14 females; mean age = 27.1, S.D. = 6.8) healthy volunteers with no history of Axis I psychiatric illness as determined from the SCID-NP (Spitzer and Williams, 1998) and 19 (14 males, 5 females; mean age = 26.3, S.D. = 5.9) patients participated in this study. All patients were derived from a larger cohort of patients participating in clinical treatment trials

of antipsychotic medications at Zucker Hillside Hospital who had both magnetic resonance (MR) imaging and genotype data available for analysis. Genotype distributions were as follows (patient, healthy volunteer): *phelphe* (2,0), *leulphe* (3,6) and *leulleu* (14,19). Diagnoses of patients were based on the Structured Clinical Interview for Axis I DSM-IV Disorders (First et al., 1994) and included schizophrenia ($N = 17$), schizoaffective ($N = 1$) or schizophreniform ($N = 1$) disorder. All patients with MR imaging data were experiencing a first-episode of illness with a median of 0 weeks exposure to antipsychotic treatment (range = 0–23 weeks). Eleven patients were antipsychotic drug-naïve at the time of the MR imaging exam. Mean age at onset of illness was 22.7 (S.D. = 5.1) years.

Classification of handedness for individuals with MR imaging data was based on a modified version of the Edinburgh Inventory consisting of 20 items (Oldfield, 1971) using the following formula: (total right – total left)/(total right + total left). Subjects with a laterality quotient greater than .70 were classified as dextral and the rest were classified as nondextral (Schachter et al., 1987). Mean (S.D.) laterality quotient for patients was .76 (S.D. = .58) and for controls was .72 (S.D. = .53). Handedness for 1 patient and 2 healthy volunteers was based on preference for handwriting alone.

Two hundred patients with a diagnosis of schizophrenia ($N = 156$) or schizoaffective disorder ($N = 44$) based on the Structured Clinical Interview for Axis I DSM-IV Disorders (First et al., 1994) had genotype data available and complete item information available from the psychosis module for analysis. Among the 200 patients 11 were *phelphe* homozygotes, 45 were *leulphe* heterozygotes and 144 were *leulleu* homozygotes.

Exclusion criteria for patients and healthy volunteers included any known genetic, neurologic or seizure disorder or meeting DSM-IV criteria for mental retardation. Additional exclusion criteria for healthy volunteers included an Axis I diagnosis. The sample was limited to Caucasians to control for potential population effects. All procedures were approved by the North Shore - Long Island Jewish Health System IRB and written informed consent was obtained from all participants.

2.2. Symptom measures

We assessed positive and negative symptoms using the psychosis module of the Structured Clinical Interview for Axis I DSM-IV Disorders (First et al., 1994). A negative symptom score was formed by averaging the avolition, alogia and affective flattening scores. The positive symptom score was formed by averaging the delusion and hallucination items. The delusion score was computed as the average of referential delusions, paranoid delusions, grandiose delusions, somatic delusions, other delusions, delusions of being controlled, thought broadcasting, and bizarre delusions. The hallucination score was computed as the average of auditory hallucinations, visual hallucinations, tactile hallucinations and other hallucinations.

2.3. Magnetic resonance (MR) imaging methods

MR imaging exams were conducted at the Long Island Jewish Medical Center in the coronal plane using a 3D Fast SPGR with IR Prep (TR = 12.7 or 14.7, TE = 4.5 or 5.5 ms, FOV = 22 cm) on a 1.5 T whole body superconducting imaging system (General Electric, Milwaukee, WI). This sequence produced 124 contiguous images (slice thickness = 1.5 mm) through the whole head with nominal in-plane resolution of .86 mm × .86 mm in a 256 × 256 matrix. All measurements were completed using the MEDx software program (MEDx, 1998). MR images were aligned along the anterior and posterior commissures for standardization across subjects and flipped randomly in the right–left axis. Scans were mixed together randomly and no identifying information was available to the operator from the scan. All measurements were thus completed by an operator who was blind to group membership, genotype status and hemisphere. All scans were reviewed by a member of the research team during acquisition and any scan with significant artifacts was repeated.

2.4. Total intracranial contents

Measurement of total intracranial contents was completed in MEDx by computing the volume of the total cerebrum, cerebrospinal fluid, cerebellum

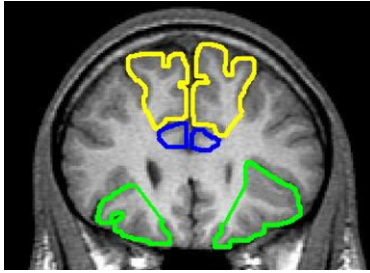


Fig. 1. Illustration of the frontal lobe regions-of-interest. Notes: superior frontal gyrus = yellow; anterior cingulate gyrus = blue; orbital frontal lobe = Green. Outlined regions were automatically segmented into gray and white matter using a thresholding method generated from gray-level histograms (see text and Otsu, 1979 for details)

and brainstem. Inter-rater reliability between two raters as assessed by intra-class correlations [ICCs] in 9 cases was .99.

2.5. Frontal lobe subregions

The frontal lobe subregions were measured in the coronal plane using methods described previously (Szeszko et al., 1999, 2004) that were adapted from Rademacher et al. (1992). An illustration of these regions at the level of the genu of the corpus callosum is provided in Fig. 1 and the limiting boundaries for these regions are illustrated in Fig. 2. This method utilizes the cerebral sulci in combination with a set of coronal planes that “close” the borders of selected regions of interest. Although the most anterior portion of these frontal regions may be excluded from measurement, the major strength of an approach based on the sulcal anatomy is to allow consistency of measurement of cytoarchitectonic regions across subjects given the fundamental problem associated with several prior schemes based on the use of invariant landmarks. The boundaries of the superior frontal gyrus were (anterior, posterior, lateral, medial): tip of the cingulate sulcus, connection of the superior and precentral sulci, superior frontal sulcus and cingulate sulcus. The boundaries of the anterior cingulate gyrus were (anterior, posterior, ventral, dorsal): tip of the cingulate sulcus, connection of the superior and precentral sulci, callosal sulcus and cingulate sulcus. Based on our empirical (Szeszko et al., 1999, 2000) and theoretical (Bilder and Szeszko, 1996; Bilder and Degreef, 1991; Christensen and Bilder,

2000) work the superior frontal gyrus and anterior cingulate gyrus volumes were summed together to form a single measure of “dorsal” brain volume to minimize potential Type I error in the analysis of the brain structure volumes.

The boundaries of the orbital frontal region, which served as the “ventral” brain volume were (anterior, posterior, lateral and medial): last appearance of the anterior horizontal ramus, last appearance of the olfactory sulcus, anterior horizontal ramus/circular sulcus of insula and the olfactory sulcus. Because one of the limiting sulci required for measurement of the orbital frontal region (i.e., the anterior horizontal ramus) was not present in every hemisphere (Szeszko et al., 1999; Ono et al., 1990), orbital frontal volumes could not be computed in the right hemisphere for 2 patients and 5 healthy volunteers and in the left hemisphere for 1 patient and 4 healthy volunteers.

All regions were outlined manually in the coronal plane on a slice by slice basis and included both gray and white matter (see Fig. 1). After outlining the frontal region of interest an operator segmented it into gray and white matter using a thresholding method generated from gray-level histograms (Otsu, 1979) as described previously (Szeszko et al., 2004; Lim et al., 1992). Given our a priori hypothesis only gray matter volumes were included in analyses. Intra-class correlations between 2 or 3 operators (number of cases ranged from 8 to 10) for the prefrontal gray matter volumes were (right hemisphere, left hemisphere): anterior cingulate gyrus gray matter (.90, .94), superior frontal gyrus gray matter (.92, .97), orbital frontal lobe gray matter (.92, .99).

2.6. Genotyping

Genotyping methods have been described in detail previously (Hodgkinson et al., 2004). Briefly, *DISC1* leu607phe genotypes were determined using a 5'-exonuclease allelic discrimination (Taqman) assay using Reference SNP ID: rs6675281 on an Applied Biosystems 7900 Analyzer (Foster City, CA). Genotyping accuracy as determined by re-genotyping one in six samples, randomly selected, produced an overall accuracy rate >99%.

2.7. Statistical procedures

All statistical tests were conducted using SAS Version 9.1 (SAS, 2002–2003). The mixed models approach for repeated measures analysis of covariance was used to investigate the relationship between leu607phe genotype and the frontal gray matter volumes in patients and healthy volunteers. When appropriate the 95% confidence intervals are presented for the difference in group means (lower to upper bound). To control Type-I error we first inves-

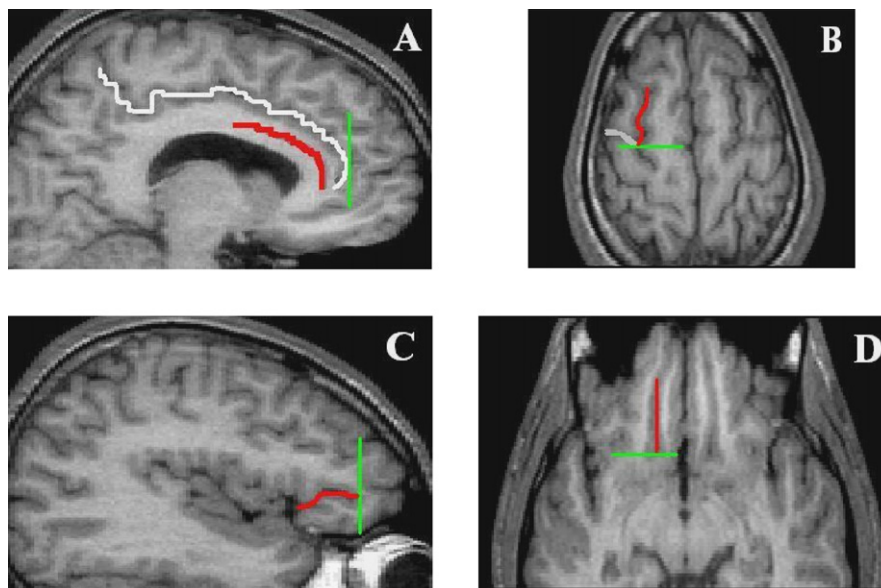


Fig. 2. Illustration of the limiting boundaries for the frontal lobe regions. Notes: Panel A, cingulate sulcus (white), callosal sulcus (red) and tip of the cingulate sulcus (green); Panel B, superior frontal sulcus (SFS; red), precentral sulcus (PRC; white) and connection of the SFS and PRC (green); Panel C, anterior horizontal ramus (ahr; red) and anterior tip of the ahr (green); Panel D, olfactory sulcus (OLS, red) and posterior tip of the OLS (green).

Table 1
Sample characteristics for symptom data analysis

	Phe carriers (<i>N</i> = 56)	leu/leu homozygotes (<i>N</i> = 144)	Test statistic	d.f.	<i>p</i> -Value
Sex (M, F)	41, 15	99, 45	$\chi^2 = .38$	1	.54
Age (years)	40.7 (9.3)	38.7 (10.9)	$t = 1.3$	198	.21
Age at onset of psychotic symptoms (years) ^a	21.4 (5.3)	21.1 (6.2)	$t = .33$	190	.74
Global assessment of functioning ^a	38.3 (15.6)	38.4 (15.2)	$t = -.03$	191	.99
Hallucination severity	1.8 (.5)	1.6 (.4)	$t = 2.6$	198	.01
Delusion severity	2.1 (.5)	2.0 (.5)	$t = 1.3$	198	.20
Avolition	1.8 (.9)	1.8 (.9)	$t = .03$	198	.98
Alogia	1.5 (.8)	1.6 (.9)	$t = -1.3$	198	.19
Affective flattening	1.7 (.8)	1.8 (.8)	$t = -.80$	198	.43

Notes: Data are presented as mean \pm S.D. in parentheses, unless otherwise indicated.

^a There were missing data for the following variables: age at onset of psychotic symptoms (3 risk carriers, 5 non-risk carriers), and global assessment of functioning (1 risk carrier, 6 non-risk carriers).

tingated all higher order interactions in the original model and retained only those that remained statistically significant in the final model, along with all main effects. In addition, to increase statistical power in the analyses we combined the small number of *phelphe* homozygotes with the *leuphe* carriers to form a single group (phe carriers). The statistical model thus included group (patient versus healthy volunteer) and genotype status (phe carriers versus leu/leu homozygotes) as between subjects factors and gray matter region (dorsal versus ventral) and hemisphere (right versus left) as within subjects factors. Statistical covariates included age, sex and total intracranial contents. Age was included as a covariate because it correlated with the brain structure volumes. Intracranial volume was included as a covariate to control for nonspecific differences in brain size among individuals. Sex was included as a covariate due to the imbalance in sex distribution between the groups.

Repeated measures analysis of variance was used to investigate the relationship between leu607phe genotype and positive and negative symptoms in patients. Given the small number of *phelphe* homozygotes and the fact that they did not differ significantly from leu/phe heterozygotes in symptoms we combined these two groups into a single group (phe carriers) for analysis. Thus, of the 200 patients 56 were classified as phe carriers and 144 were classified as leu/leu homozygotes. Sample characteristics for the two genotype groups are illustrated in Table 1. In each analysis genotype group (phe carriers versus leu/leu homozygotes) was the between subjects factor. In the negative symptom analysis the within subjects factor was negative symptom score (i.e., avolition, alogia and affective flattening). In the positive symptom analysis the within subjects factor was positive symptom score (i.e., delusions and hallucinations).

Group differences for continuous demographic variables were examined using independent groups *t* tests. Chi-square tests were used to examine differences in categorical variables. Pearson Product Moment correlations were used for investigating brain structure in relationship to symptoms. Alpha was set to .05 for all analyses. Genotype frequencies were investigated using Chi-square analysis to test for Hardy–Weinberg equilibrium.

3. Results

3.1. Magnetic resonance (MR) imaging data

Mixed models repeated measures analysis of covariance did not reveal any significant main effects of group, hemisphere, or sex. There was, however, a significant genotype-by-brain region interaction for the prefrontal cortical volumes ($F = 4.48$, d.f. = 1.38, $p = .04$), which remained significant after removal of nondextral subjects from the analysis ($F = 7.01$, d.f. = 1.31, $p = .01$). Overall, carriers of the phe allele ($N = 11$) had less gray matter in the dorsal brain region compared to leu/leu homozygotes ($N = 33$) ($t = -2.24$, d.f. = 1.38, $p = .03$; 95% CI = -261 to -3609). The genotype-by-brain region-by-

hemisphere interaction was not statistically significant indicating that the observed findings were not specific to either the right or left hemisphere. Follow-up analyses revealed that this effect was statistically significant for both the superior frontal gyrus and anterior cingulate gyrus. Specifically, carriers of the phe allele had less gray matter in the superior frontal gyrus ($t = -2.19$, d.f. = 1.38, $p = .035$; 95% CI = -6700 to -370) and anterior cingulate gyrus ($t = -2.34$, d.f. = 1.38, $p = .025$; 95% CI = -2213 to -196) compared to leu/leu homozygotes. Individual brain volumes are illustrated by genotype in Fig. 3 for the anterior cingulate gyrus and Fig. 4 for the superior frontal gyrus. There were no significant differences between the two genotype groups in orbital frontal volumes. There was a significant region-by-hemisphere interaction ($F = 6.54$, d.f. = 1.35, $p = .02$) such that overall, subjects had more gray matter in the left compared to the right orbital frontal cortex ($t = 2.81$, d.f. = 1.35, $p = .008$; 95% CI = 178 to 1002). Allelic distribution for the genotype groups in the MRI analysis did not differ significantly from Hardy–Weinberg equilibrium.

Given the main effect of *DISC1* leu607phe genotype we examined possible differences in sample characteristics between phe carriers and leu/leu homozygotes that may have influenced the observed findings. There were no significant

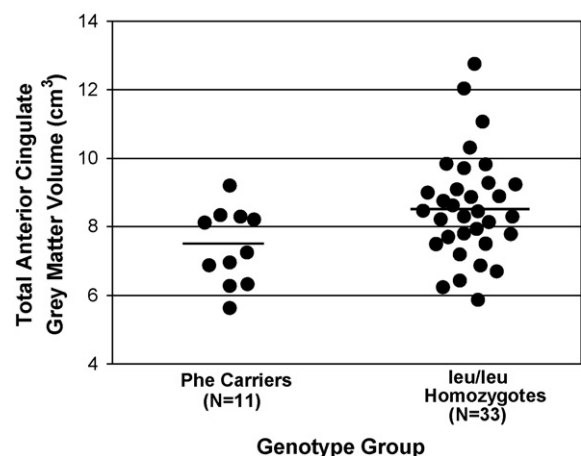


Fig. 3. Total anterior cingulate gray matter volume by leu607phe genotype. Notes: Horizontal lines represent mean values.

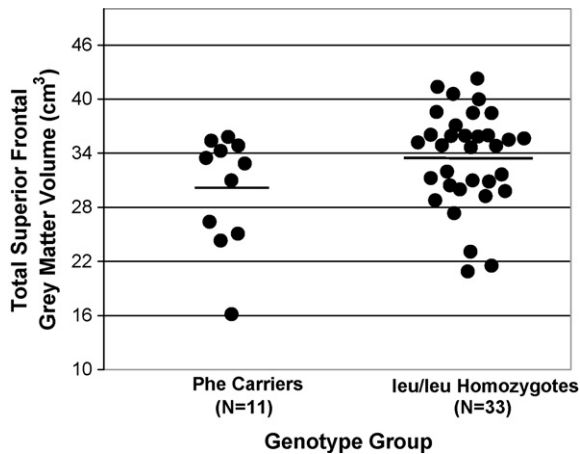


Fig. 4. Total superior frontal gray matter volume by leu607phe genotype. Notes: Horizontal lines represent mean values.

differences between the two genotype groups in distributions of sex, age, education, handedness, clinical diagnosis of schizophrenia or total intracranial contents (see Table 2). Nevertheless, we investigated the genetic hypothesis separately in patients and healthy volunteers in a post hoc analysis. Genotype accounted for a larger percentage of the variance in superior frontal gyrus volume among patients ($F = 7.59$, $d.f. = 1.14$, $p = .015$, 95% CI = $-11,142$ to -1388 ; effect size = .35) compared to healthy volunteers ($F = 3.50$, $d.f. = 1.20$, $p = .076$, 95% CI = -7288 to 2110 ; effect size = .149). Similarly, genotype accounted for a larger percentage of the variance in anterior cingulate gyrus volume among patients ($F = 5.74$, $d.f. = 1.14$, $p = .031$, 95% CI = -3082 to -170 ; effect size = .29) compared to healthy volunteers ($F = 1.32$, $d.f. = 1.20$, $p = .26$, 95% CI = -2891 to 157 ; effect size = .062).

3.2. Positive and negative symptoms

Comparison of sample characteristics for the two genotype groups in the symptom data analysis (56 phe carriers versus 144 leu/leu homozygotes) did not reveal any significant differences in distributions of age, sex, age at first psychotic symptoms and global assessment of functioning (see Table 1). Allelic

distribution for the genotype groups in the symptom data analysis did not differ significantly from Hardy–Weinberg equilibrium. ANOVA revealed a significant main effect of genotype for the positive symptom scores ($F = 5.98$, $d.f. = 1.198$, $p = .015$; 95% CI = .03 to .25). This effect was driven primarily by greater severity of hallucinations among patients who were phe carriers compared to patients who were leu/leu homozygotes ($t = 2.56$, $d.f. = 1.198$, $p = .011$; 95% CI = .04 to .32). The leu607phe polymorphism accounted for approximately 3% of the variance in hallucinations among patients. Neither the main effect of genotype for the negative symptom items nor genotype-by-negative symptom item interaction were statistically significant.

We examined the relationship between superior frontal and anterior cingulate gyrus gray matter volumes and severity of hallucinations among the subset of patients ($N = 15$) who had both MR imaging and symptom data available. These analyses revealed that less gray matter in the left superior frontal gyrus correlated significantly with increased severity of hallucinations ($r = -.53$, $d.f. = 15$, $p = .045$; Fig. 5). Neither right/left anterior cingulate gyrus gray matter nor right superior frontal gray matter volumes correlated significantly with severity of hallucinations.

4. Discussion

Our data provide evidence for an association between a *DISC1* polymorphism, leu607phe, and prefrontal cortical gray matter volume and positive symptoms. Specifically, carriers of the phe allele ($N = 11$) had less cortical gray matter volume in the superior frontal gyrus and anterior cingulate gyrus compared to leu/leu homozygotes ($N = 33$). Furthermore, in the larger cohort of patients with schizophrenia or schizoaffective disorder ($N = 200$), carriers of the phe allele ($N = 56$) had greater severity of hallucinations compared to patients who were leu/leu homozygotes ($N = 144$). Finally, we also observed a significant negative correlation between superior frontal gray matter volume and severity of hallucinations among a subgroup of 15 patients. Taken together, these results provide a potential mechanism through which *DISC1* may confer increased risk for schizophrenia or schizoaffective disorder.

Table 2
Sample characteristics for subjects with brain volume data by genotype

	Phe carriers ($N = 11$)	Leu/leu homozygotes ($N = 33$)	Statistic	d.f.	p
Sample characteristics					
Group (patients, healthy volunteers)	5/6	14/19	$\chi^2 = .03$	1	.86
Sex (male, female)	8/3	17/16	$\chi^2 = 1.51$	1	.22
Handedness (right, left)	9/2	29/4	$\chi^2 = .26$	1	.61
Age (years)	26.0 (6.8)	27.0 (6.3)	$t = -.48$	42	.64
Education (years)	14.9 (2.4)	14.4 (1.9)	$t = .76$	42	.45
Brain volume data (cm ³)					
Superior frontal gray matter	29.96 (6.23)	33.49 (5.35)			
Anterior cingulate gray matter	7.41 (1.10)	8.57 (1.54)			
Total orbital frontal gray matter	10.97 (2.80)	10.86 (2.34)			
Total intracranial volume	1351 (87)	1371 (163)			

Notes: Data are presented as mean \pm S.D. in parentheses, unless otherwise indicated.

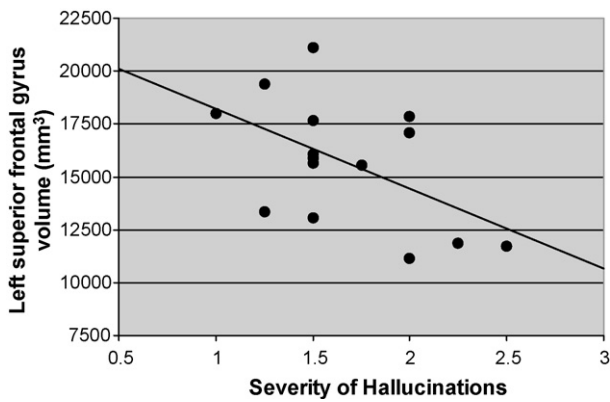


Fig. 5. Scatterplot of left superior frontal gyrus volume (mm³) and severity of hallucinations.

Despite a purported role in cortical development there are surprisingly little human data documenting an association between *DISC1* genetic variation and brain structure or function. In one study, Cannon et al. (2005) reported that a 3 locus haplotype, which included the leu607phe polymorphism, was associated with lower prefrontal gray matter density among patients with schizophrenia. Support for an association between variation in *DISC1* genotype and neurocognition was reported in two recent studies. Burdick et al. (2005) indicated that *DISC1* genotype was related to neurocognitive performance on measures of rapid visual search and verbal working memory in a cohort of 250 patients with schizophrenia. Also, Hennah et al. (2005) reported that one haplotype of *DISC1*, HEP3, was linked to short-term visual memory and attention performance in affected and unaffected offspring of patients with schizophrenia. It may therefore be noteworthy that prior associations between *DISC1* genetic variation and “working memory” performance (Burdick et al., 2005; Hennah et al., 2005) are believed to be mediated, at least in part, by gray matter in regions found in the present study to be linked to *DISC1* genetic variation in patients with schizophrenia (Rasser et al., 2005) and healthy volunteers (du Boisgueheneuc et al., 2006).

Although the functions of *DISC1* are not fully known, there is evidence that the *DISC1* protein regulates several basic developmental mechanisms in the central nervous system. The involvement of *DISC1* in neurite outgrowth was initially demonstrated by Ozeki et al. (2003) with subsequent work indicating that the *DISC1* protein participates in neurite outgrowth through interactions with FEZ1 (Miyoshi et al., 2003). Studies by Morris et al. (2003), Ozeki et al. (2003), and Brandon et al. (2004) supported this work by demonstrating a protein interaction of *DISC1* with NUDEL. *DISC1* has also been implicated in mitochondrial function and cAMP-signaling (Millar et al., 2005), which are also relevant to the pathophysiology of schizophrenia. Moreover, both animal (Schurov et al., 2004) and in vivo (Kamiya et al., 2005) studies support a role for *DISC1* in brain development, which has direct relevance for neurodevelopmental models of schizophrenia that posit cortical deficits in the pathophysiology of the disorder (Bilder and Degreef, 1991; Szeszko et al., 2003). Particularly

noteworthy in this regard are data from Kamiya et al. (2005) who reported that suppression of *DISC1* may impair migration and subsequent development of the cerebral cortex through its interactions with microtubule-associated dynein motor complex.

Our study provides evidence for an association between *DISC1* and gray matter volume in the superior frontal and anterior cingulate gyri, which have particular relevance for the neurobiology of schizophrenia. Structural alterations both in the anterior cingulate (Narr et al., 2005) and superior frontal (Jayakumar et al., 2005) gyri have been reported in patients with schizophrenia. Also, individuals at high-risk for developing schizophrenia who later became ill demonstrated abnormal anterior cingulate activity during an fMRI sentence completion task (Whalley et al., 2006). We previously suggested that the “medial frontolimbic system,” in which the frontal lobes and limbic system are linked by the cingulate bundle is isomorphic with the medial/dorsal “archicortical” system (Sanides, 1969) and can be distinguished from a ventral/lateral “paleocortical” system that comprises the olfactory cortex and peri-insular and ventral neocortices including the orbital frontal cortex. A defect in the frontolimbic system or dorsal “archicortical” trend, which encompasses both the superior frontal and anterior cingulate gyri, has been hypothesized to be critical for the stable execution of internally generated, task relevant, action-oriented behaviors (Bilder and Degreef, 1991; Bilder and Szeszko, 1996; Szeszko et al., 2002). Evidence for a genetically mediated defect in this integrated system in schizophrenia could play a role in neuropsychological deficits observed during executive functioning (Szeszko et al., 2000; Everett et al., 2001), two-choice guessing (Paulus et al., 1999), verbal fluency (Schaufelberger et al., 2005) and conflict monitoring (Kerns et al., 2005).

Our results also suggest that genetic variation in *DISC1* plays a role in positive symptoms (i.e., hallucinations) in schizophrenia. Similarly, Hennah et al. (2003) reported that variation in *DISC1* was associated with delusions, hallucinations, and negative symptoms among patients with schizophrenia. Strengthening the purported effect of *DISC1* on brain volume and its association with positive symptoms we also observed a significant inverse correlation between superior frontal gray matter volume and severity of hallucinations among patients. In that regard our results are consistent with other studies documenting an association between severity of hallucinations and prefrontal cortical abnormalities as assessed via magnetoencephalography (Kawaguchi et al., 2005) and in event related potential (Papageorgiou et al., 2005), positron emission tomography (Copolov et al., 2003) and functional magnetic resonance imaging (Lennox et al., 2000) studies in patients with schizophrenia. Moreover, other studies indicate that the prefrontal cortex may be an important region for the development of hallucinations (Castner and Goldman-Rakic, 2003) and that atypical antipsychotics that are linked with positive symptom improvement (Kane et al., 1988) alter frontal dopaminergic activity (Ichikawa and Meltzer, 1999).

There were several limitations to this study that should be acknowledged. There is the caveat of examining a single SNP

in relationship to multiple phenotypic traits, which may increase the likelihood of false positives. Thus, these findings should be replicated using larger sample sizes. In addition, the sample size for the MR imaging analysis was small, as reflected by the large confidence intervals, and thus, potentially underpowered to detect significant group differences in brain structure volumes. There is also the potential that case–control association studies may be influenced by undetected population stratification (Malhotra and Goldman, 1999). To limit this possibility, however, we limited the sample to Caucasians, and focused on a functional polymorphism that has been demonstrated previously to increase risk for schizophrenia and schizoaffective disorder and was part of a haplotype previously reported to be associated with gray matter density. Lastly, it would be important to consider potential gene–environment interactions in subsequent analyses.

In sum, these findings suggest that variation in frontal cortical gray matter volumes and positive symptoms may be explained, in part, by the leu607phe polymorphism in the *DISC1* gene. Taken together, our data provide a potential mechanism through which *DISC1* may confer increased risk for neuropsychiatric disorders such as schizophrenia or schizoaffective disorder. Future studies could focus on potential epistatic interactions of the *DISC1* gene with other genes that may increase risk for schizophrenia.

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References

- Bilder, R.M., Degreef, G., 1991. Morphologic markers of neurodevelopmental paths to schizophrenia. In: Mednick, S.A., Cannon, T.D., Barr, C.E., LaFosse, J.M. (Eds.), *Developmental Neuropathology of Schizophrenia*. Plenum Press, New York, pp. 167–190.
- Bilder, R.M., Szeszko, P.S., 1996. Structural neuroimaging and neuropsychological impairments. In: Pantellis, C., Nelson, H.E., Barnes, T.R.E. (Eds.), *The Neuropsychology of Schizophrenia*. John Wiley & Sons, Sussex UK, pp. 279–298.
- Blackwood, D.H., Fordyce, A., Walker, M.T., St Clair, D.M., Porteous, D.J., Muir, W.J., 2001. Schizophrenia and affective disorders—co-segregation with a translocation at chromosome 1q42 that directly disrupts brain-expressed genes: clinical and P300 findings in a family. *American Journal of Human Genetics* 69, 428–433.
- Blackwood, D.H., Muir, W.J., 2004. Clinical phenotypes associated with *DISC1*, a candidate gene for schizophrenia. *Neurotoxicity Research* 6, 35–41.
- Brandon, N.J., Handford, E.J., Schurov, I., Rain, J.C., Pelling, M., Duran-Jimeniz, B., et al., 2004. Disrupted in Schizophrenia 1 and Nudel form a neurodevelopmentally regulated protein complex: implications for schizophrenia and other major neurological disorders. *Molecular and Cellular Neurosciences* 25, 42–55.
- Burdick, K.E., Hodgkinson, C.A., Szeszko, P.R., Lencz, T., Ekholm, J.M., Kane, J.M., et al., 2005. *DISC1* and neurocognitive function in schizophrenia. *Neuroreport* 16, 1399–1402.
- Cannon, T.D., Hennah, W., van Erp, T.G., Thompson, P.M., Lonnqvist, J., Huttunen, M., et al., 2005. Association of *DISC1/TRAX* haplotypes with schizophrenia, reduced prefrontal gray matter, and impaired short- and long-term memory. *Archives of General Psychiatry* 62, 1205–1213.
- Cannon, T.D., Thompson, P.M., van Erp, T.G., Toga, A.W., Poutanen, V.P., Huttunen, M., et al., 2002. Cortex mapping reveals regionally specific patterns of genetic and disease-specific gray-matter deficits in twins discordant for schizophrenia. *Proceedings of the National Academy of Sciences of the United States of America* 99, 3228–3233.
- Castner, S.A., Goldman-Rakic, P.S., 2003. Amphetamine sensitization of hallucinatory-like behaviors is dependent on prefrontal cortex in nonhuman primates. *Biological Psychiatry* 54, 105–110.
- Christensen, B.K., Bilder, R.M., 2000. Dual cytoarchitectonic trends: an evolutionary model of frontal lobe functioning and its application to psychopathology. *Canadian Journal of Psychiatry* 45, 247–256.
- Coplov, D.L., Seal, M.L., Maruff, P., Ulusoy, R., Wong, M.T., Tochon-Danguy, H.J., et al., 2003. Cortical activation associated with the experience of auditory hallucinations and perception of human speech in schizophrenia: a PET correlation study. *Psychiatry Research* 122, 139–152.
- DeRosse, P., Hodgkinson, C.A., Lencz, T., Burdick, K.E., Kane, J.M., Goldman, D., Malhotra, A.K., 2007. Disrupted in schizophrenia I genotype and positive symptoms in schizophrenia. *Biological Psychology* 15, 1208–1210.
- du Boisgueheneuc, F., Levy, R., Volle, E., Seassau, M., Duffau, H., Kinkingnehun, S., et al., 2006. Functions of the left superior frontal gyrus in humans: a lesion study. *Brain* 129, 3315–3328.
- Erkwoh, R., Sabri, O., Steinmeyer, E.M., Bull, U., Sass, H., 1997. Psychopathological and SPECT findings in never-treated schizophrenia. *Acta Psychiatrica Scandinavica* 96, 51–57.
- Everett, J., Lavoie, K., Gagnon, J.F., Gosselin, N., 2001. Performance of patients with schizophrenia on the Wisconsin Card Sorting Test (WCST). *Journal of Psychiatry and Neuroscience* 26, 123–130.
- First, M.B., Spitzer, R.L., Gibbon, M., Williams, J.B.W., 1994. *Structured Clinical Interview for Axis I DSM-IV Disorders—Patient Edition (SCID-I/P)*. New York State Psychiatric Institute, New York.
- Hennah, W., Tuulio-Henriksson, A., Paunio, T., Ekelund, J., Varilo, T., Partonen, T., et al., 2005. A haplotype within the *DISC1* gene is associated with visual memory functions in families with a high density of schizophrenia. *Molecular Psychiatry* 10, 1097–1103.
- Hennah, W., Varilo, T., Kestila, M., Paunio, T., Arajärvi, R., Haukka, J., et al., 2003. Haplotype transmission analysis provides evidence of association for *DISC1* to schizophrenia and suggests sex-dependent effects. *Human Molecular Genetics* 12, 3151–3159.
- Hodgkinson, C.A., Goldman, D., Jaeger, J., Persaud, S., Kane, J.M., Lipsky, R.H., et al., 2004. Disrupted in schizophrenia 1 (*DISC1*): association with schizophrenia, schizoaffective disorder, and bipolar disorder. *American Journal of Human Genetics* 75, 862–872.
- Ichikawa, J., Meltzer, H.Y., 1999. Relationship between dopaminergic and serotonergic neuronal activity in the frontal cortex and the action of typical and atypical antipsychotic drugs. *European Archives of Psychiatry and Clinical Neuroscience* 249, 90–98.
- Jayakumar, P.N., Venkatasubramanian, G., Gangadhar, B.N., Janakiramaiah, N., Keshavan, M.S., 2005. Optimized voxel-based morphometry of gray matter volume in first-episode, antipsychotic-naïve schizophrenia. *Progress in Neuropsychopharmacology & Biological Psychiatry* 29, 587–591.
- Kamiya, A., Kubo, K., Tomoda, T., Takaki, M., Youn, R., Ozeki, Y., et al., 2005. A schizophrenia-associated mutation of *DISC1* perturbs cerebral cortex development. *Nature Cell Biology* 7, 1167–1178.
- Kane, J., Honigfeld, G., Singer, J., Meltzer, H., 1988. Clozapine for the treatment-resistant schizophrenic. A double-blind comparison with chlorpromazine. *Archives of General Psychiatry* 45, 789–796.
- Kawaguchi, S., Ukai, S., Shinosaki, K., Ishii, R., Yamamoto, M., Ogawa, A., et al., 2005. Information processing flow and neural activations in the dorsolateral prefrontal cortex in the Stroop task in schizophrenic patients: A spatially filtered MEG analysis with high temporal and spatial resolution. *Neuropsychobiology* 51, 191–203.
- Kerns, J.G., Cohen, J.D., MacDonald 3rd, A.W., Johnson, M.K., Stenger, V.A., Aizenstein, H., et al., 2005. Decreased conflict- and error-related activity in

- the anterior cingulate cortex in subjects with schizophrenia. *American Journal of Psychiatry* 162, 1833–1839.
- Lennox, B.R., Park, S.B., Medley, I., Morris, P.G., Jones, P.B., 2000. The functional anatomy of auditory hallucinations in schizophrenia. *Psychiatry Research* 100, 13–20.
- Lim, K.O., Zipursky, R.B., Watts, M.C., Pfefferbaum, A., 1992. Decreased gray matter in normal aging: an in magnetic resonance study. *Journal of Gerontology* 47, B26–B30.
- Malhotra, A.K., Goldman, D., 1999. Benefits and pitfalls encountered in psychiatric genetic association studies. *Biological Psychiatry* 45, 544–550.
- MEDx (1998): Sterling, VA: Sensor Systems.
- Millar, J.K., Christie, S., Anderson, S., Lawson, D., Hsiao-Wei Loh, D., Devon, R.S., Arveiler, B., et al., 2001. Genomic structure and localisation within a linkage hotspot of Disrupted In Schizophrenia 1, a gene disrupted by a translocation segregating with schizophrenia. *Molecular Psychiatry* 6, 173–178.
- Millar, J.K., James, R., Christie, S., Porteous, D.J., 2005. Disrupted in schizophrenia 1 (DISC1): subcellular targeting and induction of ring mitochondria. *Molecular and Cellular Neurosciences* 30, 477–484.
- Millar, J.K., Wilson-Annan, J.C., Anderson, S., Christie, S., Taylor, M.S., Semple, C.A., et al., 2000. Disruption of two novel genes by a translocation cosegregating with schizophrenia. *Human Molecular Genetics* 9, 1415–1423.
- Miyoshi, K., Honda, A., Baba, K., Taniguchi, M., Oono, K., Fujita, T., et al., 2003. Disrupted-In-Schizophrenia 1, a candidate gene for schizophrenia, participates in neurite outgrowth. *Molecular Psychiatry* 8, 685–694.
- Morris, J.A., Kandpal, G., Ma, L., Austin, C.P., 2003. DISC1 (Disrupted-In-Schizophrenia 1) is a centrosome-associated protein that interacts with MAP1A, MIPT3, ATF4/5 and NUDEL: regulation and loss of interaction with mutation. *Human Molecular Genetics* 12, 1591–1608.
- Narr, K.L., Bilder, R.M., Toga, A.W., Woods, R.P., Rex, D.E., Szeszko, P.R., et al., 2005. Mapping cortical thickness and gray matter concentration in first episode schizophrenia. *Cerebral Cortex* 15, 708–719.
- Oldfield, R.C., 1971. The assessment and analysis of handedness: The Edinburgh Inventory. *Neuropsychologia* 9, 97–114.
- Ono, M., Kubik, S., Abernath, C.D., 1990. Atlas of the Cerebral Sulci. Georg Thieme Verlag, Stuttgart.
- Otsu, N.A., 1979. Thresholding selection method from gray-level histogram. *IEEE* 9, 62–66.
- Ozeki, Y., Tomoda, T., Kleiderlein, J., Kamiya, A., Bord, L., Fujii, K., et al., 2003. Disrupted-in-Schizophrenia-1 (DISC-1): mutant truncation prevents binding to NudE-like (NUDEL) and inhibits neurite outgrowth. *Proceedings of the National Academy Science of the United States of America* 100, 289–294.
- Papageorgiou, C., Lykouras, L., Alevizos, B., Ventouras, E., Mourtzouchou, P., Uzunoglu, N., et al., 2005. Psychophysiological differences in schizophrenics with and without delusional misidentification syndromes: a P300 study. *Progress in Neuropsychopharmacology & Biological Psychiatry* 29, 593–601.
- Paulus, M.P., Geyer, M.A., Braff, D.L., 1999. Long-range correlations in choice sequences of schizophrenic patients. *Schizophrenia Research* 35, 69–75.
- Porteous, D.J., Thomson, P., Brandon, N.J., Millar, J.K., 2006. The genetics and biology of DISC1—an emerging role in psychosis and cognition. *Biological Psychiatry* 60, 123–131.
- Rademacher, J., Galaburda, A.M., Kennedy, D.N., Filipek, P.A., Caviness, V.S., 1992. Human cerebral cortex: Localization, parcellation, and morphometry with magnetic resonance imaging. *Journal of Cognitive Neuroscience* 4, 352–374.
- Rasser, P.E., Johnston, P., Lagopoulos, J., Ward, P.B., Schall, U., Thienel, R., et al., 2005. Functional MRI BOLD response to Tower of London performance of first-episode schizophrenia patients using cortical pattern matching. *Neuroimage* 26, 941–951.
- Sanfilipo, M., Lafargue, T., Rusinek, H., Arena, L., Loneragan, C., Laitin, A., et al., 2000. Volumetric measure of the frontal and temporal lobe regions in schizophrenia: relationship to negative symptoms. *Archives of General Psychiatry* 57, 471–480.
- Sanides, F., 1969. Comparative architectonics of the neocortex of mammals and their evolutionary interpretation. *Annals of the New York Academy of Sciences* 167, 404–423.
- SAS vsn 9.1: Copyright © 2002–2003 by SAS Institute Inc., Cary, NC, USA.
- Schachter, S.C., Ransil, B.J., Geschwind, N., 1987. Associations of handedness with hair color and learning disabilities. *Neuropsychologia* 25, 269–276.
- Schaufelberger, M., Senhorini, M.C., Barreiros, M.A., Amaro Jr., E., Menezes, P.R., Seazufca, M., et al., 2005. Frontal and anterior cingulate activation during overt verbal fluency in patients with first episode psychosis. *Revista Brasileira de Psiquiatria* 27, 228–232.
- Schurov, I.L., Handford, E.J., Brandon, N.J., Whiting, P.J., 2004. Expression of disrupted in schizophrenia 1 (DISC1) protein in the adult and developing mouse brain indicates its role in neurodevelopment. *Molecular Psychiatry* 9, 1100–1110.
- Spitzer, R.L., Williams, D., 1998. Structured Clinical Interview for Diagnoses-Nonpatient Version. New York State Psychiatric Institute, New York.
- St Clair, D., Blackwood, D., Muir, W., Carothers, A., Walker, M., Spowart, G., et al., 1990. Association within a family of a balanced autosomal translocation with major mental illness. *Lancet* 336, 13–16.
- Szeszko, P.R., Bilder, R.M., Lencz, T., Ashtari, M., Goldman, R.S., Reiter, G., et al., 2000. Reduced anterior cingulate gyrus volume correlates with executive dysfunction in men with first-episode schizophrenia. *Schizophrenia Research* 43, 97–108.
- Szeszko, P.R., Bilder, R.M., Lencz, T., Pollack, S., Alvir, J.M., Ashtari, M., et al., 1999. Investigation of frontal lobe subregions in first-episode schizophrenia. *Psychiatry Research: Neuroimaging* 90, 1–15.
- Szeszko, P.R., Goldberg, E., Gunduz-Bruce, H., Ashtari, M., Robinson, D., Malhotra, A.K., et al., 2003. Smaller anterior hippocampal formation volume in antipsychotic-naive patients with first-episode schizophrenia. *American Journal of Psychiatry* 160, 2190–2197.
- Szeszko, P.R., Lipsky, R., Mentschel, C., Robinson, D., Gunduz-Bruce, H., Sevy, S., et al., 2005. Brain-derived neurotrophic factor val66met polymorphism and volume of the hippocampal formation. *Molecular Psychiatry* 10, 631–636.
- Szeszko, P.R., MacMillan, S., McMeniman, M., Chen, S., Baribault, K., Lim, K.O., et al., 2004. Brain structural abnormalities in psychotropic drug-naive pediatric patients with obsessive-compulsive disorder. *American Journal of Psychiatry* 161, 1049–1056.
- Szeszko, P.R., Strous, R.D., Goldman, R.S., Ashtari, M., Knuth, K.H., Lieberman, J.A., et al., 2002. Neuropsychological correlates of hippocampal volumes in patients experiencing a first episode of schizophrenia. *American Journal of Psychiatry* 159, 217–226.
- Whalley, H.C., Simonotto, E., Moorhead, W., McIntosh, A., Marshall, I., Ebmeier, K.P., et al., 2006. Functional imaging as a predictor of schizophrenia. *Biological Psychiatry* 60, 454–462.